



AN OVERVIEW ON THERAPEUTIC ROLE OF COMBINATION OF PROBIOTICS IN TREATING PEPTIC ULCER

**Mona Motallebi^{*1}, Surya Narayan Das², Lisa Nayak³, Santoshi Gope⁴,
Smitarashmi Sinha⁵ and Yennana Jaya Vijaya⁶**

Department of Pharmacology, Gayatri College of Pharmacy, Gayatri Vihar, Jamadarpali, Sambalpur- 768200, Odisha

ARTICLE INFO

Article History:

Received 12th October, 2022

Received in revised form 23rd November, 2022

Accepted 7th December, 2022

Published online 28th January, 2023

Key words:

Peptic ulcer, lactobacillus, NSAID,
TNF- α

ABSTRACT

Peptic ulcer is a chronic gastrointestinal disorder characterised by localised erosion in the stomach lining those results in to abdominal pain, hyper acidity, bleeding, nausea, bloating or diarrhoea. The most common cause of peptic ulcer is infection due to Helicobacter pylori or prolonged use of Non-Steroidal Anti-Inflammatory (NSAID) drugs. This review aims to provide an overview on the combination of different strands of Lactic Acid Bacteria (LAB) and bifidobacterial involving mechanisms that protects the gastric mucosal layer, helps in upregulation of prostaglandin for improving stomach lining, anti-inflammatory property of probiotics in peptic ulcer caused due to pro-inflammatory cytokines.

INTRODUCTION

Peptic ulcer is an acid-induced lesion of the digestive tract that is usually located in the stomach or proximal duodenum, and is characterized by denuded mucosa with the defect extending into the submucosa or muscularis propria^[1]. The estimated prevalence of peptic ulcer disease in the general population is 5-10%^[2], but recent epidemiological studies have shown a decrease in the incidence, rates of hospital admissions, and mortality associated with peptic ulcer^[3,4]. This is most likely secondary to the introduction of new therapies and improved hygiene, which resulted in a decline in Helicobacter pylori (*H. pylori*) infections.

Traditionally, mucosal disruption in patients with the acid peptic disease is considered to be a result of a hypersecretory acidic environment together with dietary factors or stress. Risk factors for developing peptic ulcer include *H. pylori* infection, alcohol and tobacco consumption, non-steroidal anti-inflammatory drugs (NSAIDs) use, and Zollinger-Ellison syndrome^[5]. The main risk factors for both gastric and duodenal ulcers are *H. pylori* infection and NSAID use^[6]. However, only a small proportion of people affected with *H. pylori* or using NSAIDs develop peptic ulcer disease, meaning that individual susceptibility is important in the beginning of mucosal damage. Functional polymorphisms in different cytokine genes are associated with peptic ulcers. For example, polymorphisms of interleukin 1 beta (IL1B) affect mucosal interleukin 1 β production, causing *H. pylori*-associated gastroduodenal diseases^[7].

On the other hand, the risk of complications of peptic ulcer is increased four times in NSAID users, and two times in aspirin users^[8]. The concomitant use of NSAIDs or aspirin with

anticoagulants, corticosteroids, and selective serotonin reuptake inhibitors increase the risk of upper gastrointestinal bleeding^[9]. Although many people who use NSAIDs or aspirin have concurrent *H. pylori* infection, their interaction in the pathogenesis of peptic ulcer disease remains controversial. A meta-analysis of observational studies resulted in a conclusion that NSAIDs, aspirin use, and *H. pylori* infection increase the risk of peptic ulcer disease independently^[10].

H. pylori-negative, NSAID-negative, and aspirin-negative peptic ulcer disease, which is classified as an idiopathic ulcer, can be diagnosed in about one-fifth of cases^[11]. It is caused by the imbalance between factors that contribute to mucosal integrity and aggressive insults, but the pathogenic mechanisms behind the development of idiopathic peptic ulcer are still unknown^[5]. A Danish study showed that psychological stress could increase the incidence of peptic ulcer^[12]. Other etiologies include ischemia, drugs (steroids, chemotherapeutic agents) and radiotherapy, viruses, histamine, eosinophilic infiltration, gastric bypass surgery, and metabolic disturbances^[13].

Probiotics

Probiotics are defined as live microorganisms that can confer beneficial properties for health when consumed by individuals^[14,15,16]. To be considered as probiotics, microorganisms must meet several requirements, including functional and safety aspects^[17,18]. Probiotic bacteria must be non-pathogenic and should produce antibacterial substances to successfully compete with pathogens^[19,20]. They should also be tolerant to gastric acid and bile salts, and should be able to colonize and persist in the gastrointestinal epithelium^[21,22,23]. In addition, potential probiotic bacteria should meet safety aspects,

*Corresponding author: **Mona Motallebi**

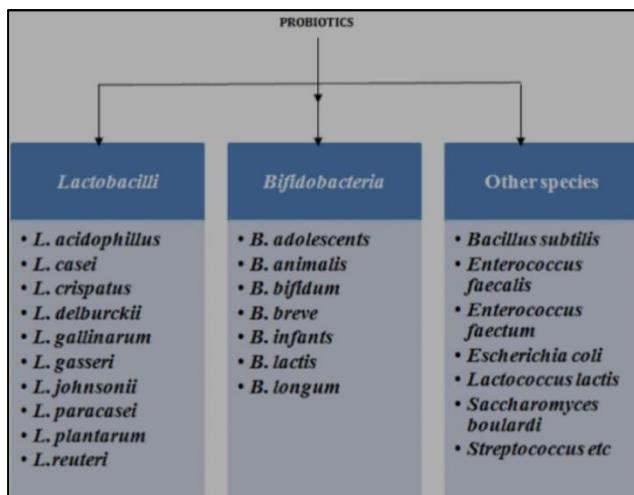
Department of Pharmacology, Gayatri College of Pharmacy, Gayatri Vihar, Jamadarpali, Sambalpur- 768200, Odisha

including the absence of transmissible antibiotic-resistance genes^[24] and the capacity to synthesize haemolysin or other toxic compounds such as nocice biogenic amines (histamine, tyramine, putrescine, cadaverine phenylethylamine or tryptamine)^[25].

Historically, the concept of probiotics began around 1900 by the Nobel laureate Elie Metchnikoff who discovered that the consumption of live bacteria (*Lactobacillus bulgaricus*) in yogurt or fermented milk improves the biological features of the gastrointestinal tract^[26,27]. The Food and Agriculture Organization and the International Scientific Association for Probiotics and Prebiotics define probiotics as live microorganisms which, when administered in adequate amounts, confer a health benefit on the host^[28]

The gut microbiota includes ~30 species of *Bifidobacterium*, 52 species of *Lactobacillus*, and others, such as *Streptococcus* and *Enterococcus*^[29]. The most extensively studied probiotics for treating and/or preventing gastrointestinal diseases are lactic acid bacteria, namely *Lactobacillus* and *Bifidobacterium* species. While these species are non-pathogenic, they can resist the harsh luminal environment of the gastrointestinal tract^[30]

Strains of Probiotics:-

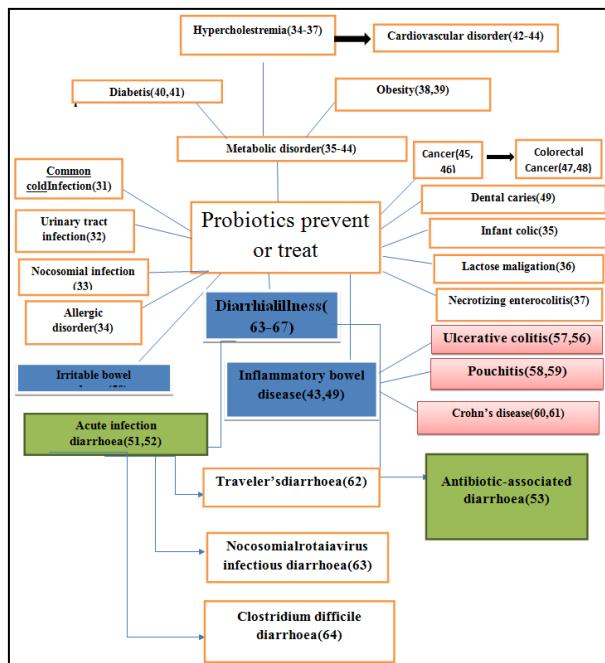


The need of Combination of Probiotics

As with all antibiotics, *H. pylori* medications often cause diarrhoea, nausea, and vomiting, which lead to poor tolerance and ultimately decreased patient compliance, the single most important factor in *H. pylori* eradication.^[65,66] Graham *et al*^[67] reported *H. pylori* eradication rates of 96% in high medication-compliant patients (taking ≥60% of the prescribed antibiotics), while only 69% eradication rates were observed among low medication-compliant patients (taking <60% of prescribed antibiotics). With the decline in *H. pylori* eradication rates, novel therapeutic alternatives are being studied and evaluated. Eradication rates are highest during the early phase of treatment when antibiotic sensitivity and patient compliance are greatest. Early treatment failure results in elevated risk of secondary antibiotic resistance due to the need for additional, less effective antibiotics used over longer periods of time with the possibility of additional medication side effects, thereby perpetuating the increase in antibiotic resistance and decreased medication compliance cycle.^[68]

Gong *et al*^[69] reported lower odds of *H. pylori* eradication with triple therapy alone, compared to triple therapy with probiotic supplementation (odds ratio [OR] 0.58; 95%

confidence interval [CI], 0.50-0.68; **P**<0.05). Significant reductions in side effects, including nausea, vomiting, bloating, epigastric pain, diarrhoea, constipation, taste distortion, and skin rash, were also observed.^[70]



Lactobacillus and *Bifidobacterium* are resistant to low pH and tolerant to a wide range of temperatures as well as can promote gut maturation and integrity, protect against pathogens, and modulate the immune system^[71]. In our paper, the study group was supplemented with live combined *Bifidobacterium*, *Lactobacillus*, and *Enterococcus* capsules, and this combination has been reported in treatment of irritable bowel syndrome^[72]. *Hp* is difficult to be treated since it acquires resistance to common antibiotics. Probiotics combined with antibiotics are currently used to eradicate *Hp*. Probiotics are useful in treating intestinal diseases such as diarrhea^[73].

It has been documented that probiotics combined with triple therapy for treating peptic ulcer infected by *Hp* can greatly improve the eradication rate of *Hp* and increase recovery rate of patients, with less adverse reaction^[74]. Several studies reveal the favorable effect of probiotics against *Hp* via different mechanisms including strength of mucosal barrier, competition for adhesion, and immunomodulatory mechanisms^[74,75].

Probiotics can also inhibit TNF-α expression, generating an immunosuppressant and anti-inflammatory effect as a response; this has been frequently reported in recent studies and is the focus of the present study.^[76]

CONCLUSION

The aim of this review was to summarize existing evidence on the mechanisms through which probiotics may exert an anti-ulcer activity and be used in case of gastric/peptic ulcer condition. The combination form of probiotics is preferable for better treatment and to prevent relapse of the disease like peptic ulcer.

Reference

- Narayanan M., Reddy K.M., Marsicano E. Peptic ulcer disease and Helicobacter pylori infection. Mo. Med. 2018;

- 115:219-224. [PMCfree article] [PubMed] [Google Scholar]
2. Lanas A., Chan F.K.L. Peptic ulcer disease. *Lancet*. 2017; 390:613-624. doi: 10.1016/S0140-6736(16)32404-7. [PubMed] [CrossRef] [Google Scholar]
 3. Lanas A., García-Rodríguez L.A., Polo-Tomás M., Ponce M., Quintero E., Perez-Aisa M.A., Gisbert J.P., Bujanda L., Castro M., Muñoz M., et al. The changing face of hospitalisation due to gastrointestinal bleeding and perforation. *Aliment. Pharmacol. Ther.* 2011; 33:585-591. doi: 10.1111/j.1365-2036.2010.04563.x. [PubMed] [CrossRef] [Google Scholar]
 4. Sonnenberg A. Review article: Historic changes of helicobacter pylori-associated diseases. *Aliment. Pharmacol. Ther.* 2013; 38:329-342. doi: 10.1111/apt.12380. [PubMed] [CrossRef] [Google Scholar]
 5. Søreide K., Thorsen K., Harrison E.M., Bingener J., Møller M.H., Ohene-Yeboah M., Søreide J.A. Perforated peptic ulcer. *Lancet*. 2015; 386:1288-1298. doi: 10.1016/S0140-6736(15)00276-7. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
 6. Zhang B.B., Li Y., Liu X.Q., Wang P.J., Yang B., Bian D.L. Association between vacA genotypes and the risk of duodenal ulcer: A meta-analysis. *Mol. Biol. Rep.* 2014; 41:7241-7254. doi: 10.1007/s11033-014-3610-y. [PubMed] [CrossRef] [Google Scholar]
 7. Datta De D., Roychoudhury S. To be or not to be: The host genetic factor and beyond in Helicobacter pylori mediated gastro-duodenal diseases. *World J. Gastroenterol.* 2015; 21:2883-2895. doi: 10.3748/wjg.v21.i10.2883. [PMC free article] [PubMed] [CrossRef] [Google Scholar]
 8. Lanas Á., Carrera-Lasfuentes P., Arguedas Y., García S., Bujanda L., Calvet X., Ponce J., Perez-Aisa Á., Castro M., Muñoz M., et al. Risk of upper and lower gastrointestinal bleeding in patients taking nonsteroidal anti-inflammatory drugs, antiplatelet agents, or anticoagulants. *Clin. Gastroenterol. Hepatol.* 2015; 13:906-912.e2.
 9. Masclee G.M., Valkhoff V.E., Coloma P.M., de Ridder M., Romio S., Schuemie M.J., Herings R., Gini R., Mazzaglia G., Picelli G., et al. Risk of upper gastrointestinal bleeding from different drug combinations. *Gastroenterology*. 2014; 147:784-792. doi: 10.1053/j.gastro.2014.06.007. [PubMed] [CrossRef] [Google Scholar]
 10. Huang J.Q., Sridhar S., Hunt R.H. Role of helicobacter pylori infection and non-steroidal anti-inflammatory drugs in peptic-ulcer disease: A meta-analysis. *Lancet*. 2002; 359:14-22. doi: 10.1016/S0140-6736(02)07273-2. [PubMed] [CrossRef] [Google Scholar]
 11. Charpignon C., Lesgourgues B., Pariente A., Nahon S., Pelaquier A., Gatineau-Saillant G., Roucayrol A.M., Courillon-Mallet A., Group de l'Observatoire National des Ulcères de l'Association Nationale des Hépatogastroentérologues des Hôpitaux Généraux (ANGH) Peptic ulcer disease: One in five is related to neither Helicobacter pylori nor aspirin/NSAID intake. *Aliment. Pharmacol. Ther.* 2013;38:946-954. doi: 10.1111/apt.12465. [PubMed] [CrossRef] [Google Scholar]
 12. Levenstein S., Rosenstock S., Jacobsen R.K., Jorgensen T. Psychological stress increases risk for peptic ulcer, regardless of Helicobacter pylori infection or use of nonsteroidal anti-inflammatory drugs. *Clin. Gastroenterol. Hepatol.* 2015; 13:498-506.e1. doi: 10.1016/j.cgh.2014.07.052. [PubMed] [CrossRef] [Google Scholar]
 13. McColl K.E. Helicobacter pylori-negative nonsteroidal anti-inflammatory drug-negative ulcer. *Gastroenterol. Clin. N. Am.* 2009; 38:353-361. doi: 10.1016/j.gtc.2009.03.004. [PubMed] [CrossRef] [Google Scholar]
 14. Sullivan A., Nord CE. Probiotics and gastrointestinal diseases. *J Intern Med* 2005; 257: 78-92. <http://dx.doi.org/10.1111/j.1365-2796.2004.01410.x>.
 15. Williams NT. Probiotics. *Am J Health Syst Pharm* 2010; 67:449-58. <http://dx.doi.org/10.2146/ajhp090168>.
 16. Mizock BA. Probiotics. *Dis Mon* 2015; 61:259-90. <http://dx.doi.org/10.1016/j.dismonth.2015.03.011>.
 17. Lee Y., Salminen S. Handbook of probiotics and prebiotics. 2th ed. New Jersey, USA: Johns Wiley & Sons; 2009 14-5. <http://dx.doi.org/10.1002/9780470432624>.
 18. Nuraida L. A review: Health promoting lactic acid bacteria in traditional Indonesian fermented foods. *Food Sci Hum Wellness* 2015; 4:47-55. <http://dx.doi.org/10.1016/j.fshw.2015.06.001>.
 19. Vitor J., Vale F. Alternative therapies for Helicobacter pylori: Probiotics and phytomedicine. *FEMS Immunol Med Microbiol* 2015; 63:153-64. <http://dx.doi.org/10.1111/j.1574-695X.2011.00865.x>.
 20. Butel MJ. Probiotics, gut microbiota and health. *Med Mal Infect* 2014; 44:1-8. <http://dx.doi.org/10.1016/j.medmal.2013.10.002>.
 21. Peres CM, Alves M, Hernandez-Mendoza A, Moreira E, Silva S, Bronze MR, et al. Novel isolates of lactobacilli from fermented Portuguese olive as potential probiotics. *LWT-Food Sci Technol* 2014; 59:234-46. <http://dx.doi.org/10.1016/j.lwt.2014.03.003>.
 22. Zhou JS, Pillidge CJ, Gopal PK, Gill HS. Antibiotic susceptibility profiles of new probiotic *Lactobacillus* and *Bifidobacterium* strains. *Int J Food Microbiol* 2005; 98:211-7. <http://dx.doi.org/10.1016/j.ijfoodmicro.2004.05.011>.
 23. Patel PJ, Singh SK, Panaich S, Cardozo L. The aging gut and the role of prebiotics, probiotics, and symbiotics: A review. *J Clin Gerontol Geriatr* 2014; 5:3-6. <http://dx.doi.org/10.1016/j.jcg.2013.08.003>.
 24. Coppola R, Succi M, Tremonte P, Reale A, Salzano G, Sorrentino E. Antibiotic susceptibility of *Lactobacillus rhamnosus* strains isolated from Parmigiano Reggiano cheese. *Lait* 2005; 85:193-204. <http://dx.doi.org/10.1051/lait.2005007>.
 25. Ancin-Azpilicueta C, González-Marco A, Jiménez-Moreno N. Current knowledge about the presence of

- amines in wine. *Crit Rev Food Sci Nutr* 2008; 48:257-75. <http://dx.doi.org/10.1080/10408390701289441>.
26. Podolsky SH: Metchnikoff and the microbiome: *Lancet* 380: 1810-1811, 2012.
 27. Lilly DM and Stillwell RH: Probiotics: Growth-promoting factors produced by microorganisms. *Science* 147: 747-748, 1965.
 28. Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, Morelli L, Canani RB, Flint HJ, Salminen S, et al: Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol* 11: 506-514, 2014.
 29. Wallace TC, Guarner F, Madsen K, Cabana MD, Gibson G, Hentges E and Sanders ME: Human gut microbiota and its relationship to health and disease. *Nutr Rev* 69: 392-403, 2011.
 30. Bezkorovainy A: Probiotics: Determinants of survival and growth in the gut. *Am J Clin Nutr* 73 (Suppl 2): S399-S405, 2001.
 31. Russo F, Linsalata M and Orlando A: Probiotics against neoplastic transformation of gastric mucosa: Effects on cell proliferation and polyamine metabolism. *World J Gastroenterol* 20: 13258-13272, 2014.
 32. Chen ZF, Ai LY, Wang JL, Ren LL, Yu YN, Xu J, Chen HY, Yu J, Li M, Qin WX, et al: Probiotics Clostridium butyricum and Bacillus subtilis ameliorate intestinal tumorigenesis. *Future Microbiol* 10: 1433-1445, 2015.
 33. Zhang M, Fan X, Fang B, Zhu C, Zhu J and Ren F: Effects of Lactobacillus salivarius Ren on cancer prevention and intestinal microbiota in 1,2-dimethylhydrazine-induced rat model. *J Microbiol* 53: 398-405, 2015.
 34. Sobhani I, Amiot A, Le Baleur Y, Levy M, Auriault ML, Van Nhieu JT and Delchier JC: Microbial dysbiosis and colon carcinogenesis: Could colon cancer be considered a bacteria-related disease? *Therap Adv Gastroenterol* 6: 215-229, 2013.
 35. Lankaputhra WEV and Shah NP: Survival of Lactobacillus acidophilus and Bifidobacterium in the presence of acid and bile salts. *Cult Dairy Prod* 30: 2-7, 1995.
 36. Costello M: Probiotic foods. In: Foodpro-93: International Food Processing Machinery and Technology Exhibition and Conference, Sydney Convention & Exhibition Centre, Australia, July 12-14 1993.
 37. ClarkPA, CottonLN and Martin JH: Selection of Bifidobacterium spp. for use as dietary adjuncts in cultured dairy foods: II. Tolerance to simulated pH of human stomachs. *Cult Dairy Prod*.
 38. Jacobsen CN, Rosenfeldt Nielsen V, Hayford AE, Møller PL, Michaelsen KF, Paerregaard A, Sandström B, Tvede M and Jakobsen M: Screening of probiotic activities of forty-seven strains of Lactobacillus spp. by in vitro techniques and evaluation of the colonization ability of five selected strains in humans. *Appl Environ Microbiol* 11: 4949-4956, 1999.
 39. Lick S, Drescher K and Heller JK: Survival of Lactobacillus delbrueckii subsp. bulgaricus and Streptococcus thermophilus in the terminal ileum of fistulated Göttingen minipigs. *Appl Environ Microbiol* 67: 4137-4143, 2001.
 40. Cotter PD and Hill C: Surviving the acid test: Responses of gram-positive bacteria to low pH. *Microbiol Mol Biol Rev* 67: 429-453, 2003.
 41. Fortier LC, Tourdot-Maréchal R, Diviès C, Lee BH and Guzzo J: Induction of Oenococcusoeni H+-ATPase activity and mRNA transcription under acidic conditions. *FEMS Microbiol Lett* 222: 165-169, 2003.
 42. Rius N, Solé M, Francis A and Lorén JG: Buffering capacity and membrane H+ conductance of lactic acid bacteria. *FEMS Microbiol Lett* 120: 291-296, 1994.
 43. Charalampopoulos D, Pandiella SS and Webb C: Evaluation of the effect of malt, wheat and barley extracts on the viability of potentially probiotic lactic acid bacteria under acidic conditions. *Int J Food Microbiol* 82: 133-141, 2003.
 44. Corcoran BM, Stanton C, Fitzgerald GF and Ross RP: Survival of probiotic lactobacilli in acidic environments is enhanced in the presence metabolizable sugars. *Appl Environ Microbiol* 71: 3060-3067, 2005.
 45. Haghshenas B, Abdullah N, Nami Y, Radiah D, Rosli R and YariKhosroushahi A: Microencapsulation of probiotic bacteria Lactobacillus plantarum 15HN using alginate-psyllium-fenugreek polymeric blends. *J ApplMicrobiol* 118: 1048-1057, 2015.
 46. Chen S, Zhao Q, Ferguson LR, Shu Q, Weir I and Garg S: Development of a novel probiotic delivery system based on microencapsulation with protectants. *ApplMicrobiolBiotechnol* 93: 1447-1457, 2012.
 47. Kailasapathy K: Microencapsulation of probiotic bacteria: Technology and potential applications. *Curr Issues IntestMicrobiol* 3: 39-48, 2002.
 48. Villena MJ, Lara-Villoslada F, Martínez MA and Hernández ME: Development of gastro-resistant tablets for the protection and intestinal delivery of Lactobacillus fermentum CECT 5716. *Int J Pharm* 487: 314-319, 2015.
 49. Rashid SK, Idris-Khodja N, Auger C, Alhosin M, Boehm N, Oswald-Mammosser M and Schini-Kerth VB: Probiotics (VSL#3) prevent endothelial dysfunction in rats with portal hypertension: Role of the angiotensin system. *PLoS One* 9: e97458, 2014.
 50. Aihara K, Kajimoto O, Hirata H, Takahashi R and Nakamura Y: Effect of powdered fermented milk with Lactobacillus helveticus on subjects with high-normal blood pressure or mild hypertension. *J Am Coll Nutr* 24: 257-265, 2005.
 51. Huang Y, Wang J, Quan G, Wang X, Yang L and Zhong L: Lactobacillus acidophilus ATCC 4356 prevents atherosclerosis via inhibition of intestinal cholesterol absorption in apolipoprotein E-knockout mice. *Appl Environ Microbiol* 80: 7496-7504, 201

52. Moroti C, Souza Magri LF, de Rezende Costa M, Cavallini DC and Sivieri K: Effect of the consumption of a new symbiotic shake on glycemia and cholesterol levels in elderly people with type 2 diabetes mellitus. *Lipids Health Dis* 11: 29, 2012.
53. Tsai TY, Chen LY and Pan TM: Effect of probiotic-fermented, genetically modified soy milk on hypercholesterolemia in hamsters. *J Microbiol Immunol Infect* 47: 1-8, 2014.
54. Pereira DI and Gibson GR: Cholesterol assimilation by lactic acid bacteria and bifidobacteria isolated from the human gut. *Appl Environ Microbiol* 68: 4689-4693, 2002.
55. Kadooka Y, Sato M, Imaizumi K, Ogawa A, Ikuyama K, Akai Y, Okano M, Kagoshima M and Tsuchida T: Regulation of abdominal adiposity by probiotics (*Lactobacillus gasseri* SBT2055) in adults with obese tendencies in a randomized controlled trial. *Eur J Clin Nutr* 64: 636-643, 2010.
56. Yoda K, Sun X, Kawase M, Kubota A, Miyazawa K, Harata G, Hosoda M, Hiramatsu M, He F and Zemel MB: A combination of probiotics and whey proteins enhances anti-obesity effects of calcium and dairy products during nutritional energy restriction in aP2-agouti transgenic mice. *Br J Nutr* 113: 1689-1696, 2015.
57. Kim SH, Huh CS, Choi ID, Jeong JW, Ku HK, Ra JH, Kim TY, Kim GB, Sim JH and Ahn YT: The anti-diabetic activity of *Bifidobacterium lactis* HY8101 in vitro and in vivo. *J ApplMicrobiol* 117: 834-845, 2014.
58. Marin ML, Lee JH, Murtha J, Ustunol Z and Pestka JJ: Differential cytokine production in clonal macrophage and T-cell lines cultured with bifidobacteria. *J Dairy Sci* 80: 2713-2720, 1997.
59. Wagner RD, Pierson C, Warner T, Dohnalek M, Hilty M and Balish E: Probiotic effects of feeding heat-killed *Lactobacillus acidophilus* and *Lactobacillus casei* to *Candida albicans*-colonized immunodeficient mice. *J Food Prot* 63: 638-644, 2000.
60. Rachmilewitz D, Kataura K, Karmeli F, Hayashi T, Reinus C, Rudensky B, Akira S, Takeda K, Lee J, Takabayashi K and Raz E: Toll-like receptor 9 signaling mediates the anti-inflammatory effects of probiotics in murine experimental colitis. *Gastroenterology* 126: 520-528, 2004.
61. Orlando A, Refolo MG, Messa C, Amati L, Lavermicocca P, Guerra V and Russo F: Antiproliferative and proapoptotic effects of viable or heat-killed *Lactobacillus paracasei* IMPC2.1 and *Lactobacillus rhamnosus* GG in HGC-27 gastric and DLD-1 colon cell lines. *Nutr Cancer* 64: 1103-1111, 2012.
62. Sanders ME, Guarner F, Guerrant R, Holt PR, Quigley EM, Sartor RB, Sherman PM and Mayer EA: An update on the use and investigation of probiotics in health and disease. *Gut* 62: 787-796, 2013.
63. Marchesi JR, Adams DH, Fava F, Hermes GD, Hirschfield GM, Hold G, Quraishi MN, Kinross J, Smidt H, Tuohy KM, et al: The gut microbiota and host health: A new clinical frontier. *Gut* 65: 330-339, 2016.
64. Kumar M, Nagpal R, Kumar R, Hemalatha R, Verma V, Kumar A, Chakraborty C, Singh B, Marotta F, Jain S and Yadav H: Cholesterol-lowering probiotics as potential biotherapeutics for metabolic diseases. *Exp Diabetes Res* 2012.
65. Ruggiero P: Use of probiotics in the fight against *Helicobacter pylori*. *World J Gastrointest Pathophysiol*. 2014;5(4):384-391. [PMC free article] [PubMed] [Google Scholar]
66. O'Connor JP, Taneike I, O'Morain C: Improving compliance with *Helicobacter pylori* eradication therapy: when and how? *Therap Adv Gastroenterol*. 2009;2(5):273-279. [PMC free article] [PubMed] [Google Scholar]
67. Graham DY, Lew GM, Malaty HM, et al: Factors influencing the eradication of *Helicobacter pylori* with triple therapy. *Gastroenterology*. 1992;102(2):493-496. [PubMed] [Google Scholar]
68. Navarro-Rodriguez T, Silva FM, Barbuti RC, et al: Association of a probiotic to a *Helicobacter pylori* eradication regimen does not increase efficacy or decreases the adverse effects of the treatment: a prospective, randomized, double-blind, placebo-controlled study. *BMC Gastroenterol*. 2013;13:56. [PMC free article] [PubMed] [Google Scholar]
69. Gong Y, Li Y, Sun Q: Probiotics improve efficacy and tolerability of triple therapy to eradicate *Helicobacter pylori*: a meta-analysis of randomized controlled trials. *Int J ClinExp Med*. 2015;8(4):6530-6543. [PMC free article] [PubMed] [Google Scholar]
70. L. Rodes, A. Khan, A. Paul et al., "Effect of probiotics *lactobacillus* and *Bifidobacterium* on gut-derived lipopolysaccharides and inflammatory cytokines: an in vitro study using a human colonic microbiota model," *Journal of Microbiology and Biotechnology*, vol. 23, no. 4, pp. 518-526, 2013. View at: Publisher Site | Google Scholar
71. L. Rodes, A. Khan, A. Paul et al., "Effect of probiotics *lactobacillus* and *Bifidobacterium* on gut-derived lipopolysaccharides and inflammatory cytokines: an in vitro study using a human colonic microbiota model," *Journal of Microbiology and Biotechnology*, vol. 23, no. 4, pp. 518-526, 2013. View at: Publisher Site | Google Scholar
72. Y. Aiba, Y. Nakano, Y. Koga, K. Takahashi, and Y. Komatsu, "A highly acid-resistant novel strain of *Lactobacillus johnsonii* no. 1088 has antibacterial activity, including that against *Helicobacter pylori*, and inhibits gastrin-mediated acid production in mice," *Microbiology*, vol. 4, no. 3, pp. 465-474, 2015. View at: Publisher Site | Google Scholar
73. Y. Aiba, Y. Nakano, Y. Koga, K. Takahashi, and Y. Komatsu, "A highly acid-resistant novel strain of *Lactobacillus johnsonii* no. 1088 has antibacterial activity, including that against *Helicobacter pylori*, and inhibits gastrin-mediated acid production in mice," *Microbiology*, vol. 4, no. 3, pp. 465-474, 2015. View at: Publisher Site | Google Scholar
74. Y. Aiba, Y. Nakano, Y. Koga, K. Takahashi, and Y. Komatsu, "A highly acid-resistant novel strain of

- Lactobacillus johnsonii no. 1088 has antibacterial activity, including that against Helicobacter pylori, and inhibits gastrin-mediated acid production in mice,” *Microbiology*, vol. 4, no. 3, pp. 465-474, 2015. View at: Publisher Site | Google Scholar
75. V. Papastergiou, S. D. Georgopoulos, and S. Karatapanis, “Treatment of helicobacter pylori infection: past, present and future,” *World Journal of Gastrointestinal Pathophysiology*, vol. 5, no. 4, pp. 392-399, 2014. View at: Publisher Site | Google Scholar
76. H. Wang, S. Li, H. Li, F. Du, J. Guan, Y. Wu, Mechanism of probiotic VSL#3 inhibiting NF- κ B and TNF- α on colitis through TLR4-NF- κ B signal pathway, *Iran, J. Public Health* 48 (2019) 1292-1300, <https://doi.org/10.18502/ijph.v48i7.2953>.
