



BACTERIOLOGICAL PROFILE AND PHENOTYPIC SIGNS OF FOOD-PROCESSING MICROBIOTA

Dudchik Natalia V., Sychik Sergei I and Nezhvinskaya Olga E

Laboratory of Microbiology, Republican Unitary Enterprise "The Scientific and Practical Centre of Hygiene", Minsk, Republic of Belarus

ARTICLE INFO

Article History:

Received 15th January, 2019

Received in revised form 7th February, 2019

Accepted 13th March, 2019

Published online 28th April, 2019

Key words:

microorganisms, contamination; biomarkers; biofilm formation, recovery methods, microbial risk analysis

ABSTRACT

Objectives: This study was undertaken to isolate and investigate bacteriological profile and phenotypic signs of food-processing microbiota strains including to *Escherichia*, *Klebsiella*, *Enterobacter*, *Staphylococcus*, *Pseudomonas*, *Citrobacter* and *Serratia* genera. Strains were isolated during hygienic monitoring of two milk-processing factories for the period from December 2013 to November 2014.

Methods: Swabbing was done in field studies according to food safety management protocols in industry for enumeration of contamination by microorganisms and isolation of strains of microbiota. Identification of microorganism was done with culture methods followed by PCR verification. Phenotypic features were studied in vitro with standard biochemical and microbiological methods according to good laboratory practice.

Results: Bacterial isolates included to *Escherichia*, *Klebsiella*, *Enterobacter*, *Citrobacter*, *Serratia*, *Staphylococcus* and *Pseudomonas* genera. Gram-positive cocci account for 20.3 % of cases of contamination. In 55 (74.3 %) cases, the pathogens belonged to the family Enterobacteriaceae. Isolate investigations included ability to hemolysis, anti-interferon, anti-lysozyme activity, lecithinase activity, ability to form biofilms in a monoculture of microorganisms. Out of 74 isolates 60 (81.1 %) showed variability of morphological and tinctorial signs.

Conclusions: Gram-negative pathogens and more specifically *Escherichia coli* account for a substantial part of bacterial milk-processing microbiota. Most isolates possessed signs of aggression and ability of biofilm formation. The obtained experimental data will be used for identification of danger factor according to the concept of the quantitative analysis of microbiological risk of food productions.

Copyright © 2019 Dudchik Natalia V et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Ensuring microbiological safety of foodstuff is one of priority tasks and directed to public health care. According to a number of epidemiologists and microbiologists, phenomenon of emergent bacterial infections with food way of transmission can be explained with change of phenotypical signs, including the ecological and pathogenetic properties of earlier studied pathogens under the influence of the amplifying anthropogenic factors. At the same time foodstuff and food-processing are represented as new environment or an ecological niche favorable for formation of new phenotypical factors of pathogenicity. Experimental studying of physiology and biochemical properties of isolates of technological environment as a result of hygienic monitoring on the basis of the algorithm offered was the purpose of the study.

MATERIALS AND METHODS

Methods for recovering microorganisms from solid surfaces. As conventional swabbing is the recommended

method¹, it is commonly used and practically applied in field studies or food safety management protocols in industry.

Methods to analyze microorganisms. Swab samples were investigated using culture methods supplemented by PCR verification². Visual identification of colonies on the selective agar were used for primary identification and further verification of isolate identification was provided by real time PCR with common laboratory procedure. Hemolytic, lecithinase activity and persistence were studied by standard methods^{3, 4, 5, 6, 7}. The study of the ability to film formation was carried out according to culture plate method with optical detection^{8, 9, 10}.

RESULTS

85 bacterial strains were isolated and studied which belong to genera *Escherichia*, *Klebsiella*, *Enterobacter*, *Staphylococcus*, *Pseudomonas*, *Citrobacter*. Apart from bacterial pathogens, 60 cases were observed to be contaminated with *Candida*, *Penicillium* and *Aspergillus* genera (data not shown). Experimental data of various isolates were depicted in table 1.

*Corresponding author: Dudchik Natalia V

Laboratory of Microbiology, Republican Unitary Enterprise "The Scientific and Practical Centre of Hygiene", Minsk, Republic of Belarus

Table 1 Phenotypical signs of Bacterial isolates

Genera	Phenotypical signs					Morphological and tinctorial properties
	hemolysis	antilysozyme activity	antiinterferon activity	lecithinase activity	biofilm forming	
<i>Escherichia</i> (19 isolates)	γ	+/-	+/-	+/-	+/-	variable
<i>Klebsiella</i> (9 isolates)	γ	+/-	+/-	+/-	max	variable
<i>Serratia</i> (10 isolates)	γ	+/-	+/-	+/-	+/-	variable
<i>Enterobacter</i> (9 isolates)	γ	+/-	+/-	+/-	+/-	variable
<i>Citrobacter</i> (8 isolates)	γ	+/-	+/-	+/-	+/-	variable
<i>Staphylococcus</i> (15 isolates)	α/β	+/-	+/-	+/-	min	variable/stable
<i>Pseudomonas</i> . (14 isolates)	α	+/-	+/-	+/-	+/-	variable/stable

β-hemolytic activity was found in all studied strains of *Staphylococcus aureus* and both α- and β-hemolytic activity was demonstrated of *Pseudomonas aeruginosa* isolates. A number of *Staphylococcus* spp. with an incomplete hemolytic phenotype have been found in our study. Isolates belong to Enterobacteriaceae family demonstrated γ-hemolytic activity. 100 % of studied strains of *Staphylococcus aureus* display lecithinase activity.

Anti-interferon and anti-lysozyme activity as persistence factors aimed at inactivating host defense mechanisms have been studied in Staphylococci. Of the 9 studied strains of *Staphylococcus aureus*, 8 display anti-lysozyme activity at a lysozyme concentration of 4 μg / ml or less, 5 strains had anti-interferon activity (interferon concentration 2 units), 3 strains had anti-interferon activity (interferon concentration 1 units). The phenomenon is accordance with good ability of staphylococcal strains grow on nutrient agar with fuzidin concentration of 0.00015-0.0003 g/l.

The studied strains have a different ability to biofilm formation in a monoculture of microorganisms. All strains of *Klebsiella pneumoniae* have a high degree of film-forming ability according to the Stepanovic criterion. Coagulase-negative staphylococcus (*S. haemolyticus*, *S. sciuri*, *S. epidermidis*) possessed minimal film-forming ability. Out of 85 isolates 60 poses variability of morphological and tinctorial signs.

DISCUSSION

It is important to note that the correct choice of methods to isolate and investigate phenotypical properties of bacterial strains¹ is significant. The choice of the recovery method has to be suitable for the type and size of the surface tested for microbiological analysis.

The phenomena of variability of morphological and tinctorial properties of isolates have been noted^{11, 12, 13}. Isolates of hospital environments as well as food-processing display atypical signs, such as strong ability to hemolysis, anti-interferon, anti-lysozyme activity, lecithinase activity, ability to form biofilms. On the other hand, recently a number of strains belonging to a class of *S. aureus* with an incomplete hemolytic phenotype.

In our study, out of 74 isolates 60 (81.1 %) poses variability of morphological and tinctorial signs. Bacteria genera *Escherichia*, *Klebsiella*, *Enterobacter*, *Staphylococcus*, *Pseudomonas*, *Citrobacter* and *Serratia* are widespread contaminants isolated during food-processing, which can cause both hospital and community-acquired infections¹³. It was reported that a lot of microbial contaminants can survive

in a range of solid surfaces, such as plastic, stainless steel, glass, and wood¹. These surfaces are subject to contamination by microorganisms responsible for the cross-contamination of food by contact with working surfaces. This preventive approach has resulted in the use of microbiological analyses of surfaces as one of the tools to control the hygiene of products and limit contamination risks. The virulence of microbial contaminants is closely associated with a variety of secreted enzymes and toxins produced by the bacteria. Recent studies have demonstrated that hemolysin also participates in the formation of the *S. aureus* biofilm¹³.

CONCLUSION

The obtained experimental data will be used for identification of danger within the concept of the quantitative analysis of microbiological risk of food productions. The increasing prevalence of strains with atypical signs within community environments further increases the dangers of microbial contaminants.

References

1. Rached Ismail, Florence Aviat, Valérie Michel, Isabelle Le Bayon, Perrine Gay-Perret, Magdalena Kutnik, Michel Fédérighi. Methods for recovering microorganisms from solid surfaces used in the food industry: a review of the literature. Int J Environ Res Public Health. 2013; 10(11): 6169–6183. doi: 10.3390/ijerph10116169.
2. Microbiology of Food and Animal Feeding Stuffs—Horizontal Methods for Sampling Techniques from Surfaces Using Contact Plates and Swabs. ISO; Geneva, Switzerland: 2004. (ISO 18593:2004).
3. Manual of Methods for General Bacteriology. Ed. P. Gerhardt, American Society for Microbiology: Washington, D.C. American Society for Microbiology, 1981.
4. Brauner A., Fridman O., Balaban N.Q. *et al.* Distinguishing between resistance, tolerance and persistence to antibiotic treatment. Nat. Rev. Microbiol. 2016, 14: 320-330.
5. Ximenes E, Hoagland L, Ku S, Li X, Ladisch M. Human pathogens in plant biofilms: Formation, physiology, and detection. Biotechnol Bioeng. 2017, 114(7), 1403-1418. doi: 10.1002/bit.26247.
6. Bukharin OV, Valyshev AV. [Microbial inhibitors of lysozyme]. Zh Mikrobiol Epidemiol Immunobiol. 2006, 4, 8-13. [Article in Russian]
7. Callewaert L, Van Herreweghe JM, Vanderkelen L, Leysen S, Voet A, Michiels CW. Guards of the great wall: bacterial lysozyme inhibitors. Trends Microbiol. 2012, 20(10), 501-10. doi: 10.1016/j.tim.2012.06.005.
8. Bodur T., Cagri-Mehmetoglu A. Removal of *Listeria monocytogenes*, *Staphylococcus aureus* and *Escherichia coli* O157:H7 biofilms on stainless steel using scallop shell powder. Food Control. 2012, 25, 1–9. doi: 10.1016/j.foodcont.2011.09.032.
9. A. Hassan [et al.] Evaluation of different methods of biofilm formation in the clinical isolates. Brazil. J. of Infectious Diseases. 2011, 15 (4), 305–311.
10. T. Mathur [at al.] Detection of biofilm formation among the clinical isolates of staphylococci: an evaluation of three different screening methods. Ind. J of Med. Microbiol. 2006, 24 (1), 25-29.

11. Haifang Zhang, Yi Zheng, Huasheng Gao, Ping Xu, Min Wang, Aiqing Li, Minhui Miao, Xiaofang Xie, Yimai Deng, Huiqin Zhou, Hong Du. identification and characterization of *Staphylococcus aureus* strains with an incomplete hemolytic phenotype. *Front Cell Infect Microbiol.* 2016, 6, 146.doi: 10.3389/fcimb.2016.00146.
12. Callewaert L, Van Herreweghe JM, Vanderkelen L, Leysen S, Voet A, Michiels CW. Guards of the great wall: bacterial lysozyme inhibitors. *Trends Microbiol.* 2012, 10, 501-510. doi: 10.1016/j.tim.2012.06.005.
13. den Reijer P. M., Haisma E. M., Lemmens-den Toom N. A., Willemse J., Koning R. A., Demmers J. A., *et al.* Detection of alpha-toxin and other virulence factors in biofilms of *Staphylococcus aureus* on polystyrene and a human epidermal model. *PLoS ONE.* 2016. 11:e0145722. 10.1371/journal.pone.0145722.

How to cite this article:

Dudchik Natalia V., Sychik Sergei I and Nezhvinskaya Olga E (2019) ' Bacteriological Profile and Phenotypic Signs of Food-Processing Microbiota', *International Journal of Current Medical And Pharmaceutical Research*, 05(04), pp 4131-4133.
