



HEMOGLOBIN FRACTIONATION BY HPLC

Gireesh. V. Achalkar*

Department of Pathology, Raichur Institute of Medical Sciences, Raichur, Karnataka-584102

ARTICLE INFO

Article History:

Received 22nd May, 2018

Received in revised form 5th

June, 2018

Accepted 16th July, 2018

Published online 28th August, 2018

Key words:

High Performance Liquid
Chromatography (HPLC),
Hemoglobin, HbA, HbA1, HbA2, HbF

ABSTRACT

Introduction: High performance liquid chromatography (HPLC) is a new technique which has demonstrated advantages in terms of accuracy and precision over conventional procedures employed in newborn and adult hemoglobinopathy screening programs for the identification of Hb variants. It has prompted the need to reassess our knowledge of hemoglobin fractions reference ranges as it relates to HPLC HB quantitation. **Materials and Methods:** Fifty microliter aliquots of fresh whole blood samples collected in ethylenediamine tetraacetic acid (EDTA) containing vacutainer tubes from normal newborn and adult individuals being screened at the were treated with a hemolyzate reagent, centrifuged at 12,000 rpm at 4°C for 15 minutes and passed through a 0.25 to 0.45 diameter syringe filter unit into a 1.5 ml tube. **Results:** In this study, the HPLC hemoglobin reference ranges derived from 100 normal adults (ages 18-50) and infants of upto 300 days of age were studied. And HPLC were chosen to establish the hemoglobin reference ranges.. The percentage values are as follows: Hb A mean of 92.3 percent; HbA1 mean 3.6 percent; Hb F mean of 3.2 percent; Hb A2 mean of 2.1 percent, respectively.

Conclusion: Application of the HPLC ranges to confront other abnormalities will prove most useful during blood screening processes. The HPLC hemoglobin ranges for normal adult and newborns have been established.

Copyright © 2018 Gireesh. V. Achalkar. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Identification of human hemoglobin variants in blood from adults and new born babies are usually carried out by cellulose acetate and citrate agar electrophoresis and also by isoelectric focusing (IEF)¹. In fact, these procedures are routinely employed in screening programs to detect sickle cell, thalassemia, and other hemoglobinopathies in affected individuals. Although, it is recognized that when used in combination, these techniques are capable of separating common hemoglobin variants like A, F, S, A2, Hb C, and Hb E; other variants are difficult to identify since they migrate together or very close. Quantification of the Hb variants by densitometry scanning of the gels is not recommended because of its sensitivity limits and inaccuracy. These limitations have prompted the use of a more versatile tool like high performance liquid chromatography (HPLC) not only to identify but to quantify a number of different and rare hemoglobins. The literature abounds with descriptive information about the use of HPLC, and others closely related like anion-exchange and reverse-phase chromatography to identify and characterize rare hemoglobin types. Reports on the identification and hemoglobin separation from newborn patients with severe alpha-thalassemia like Hb Bart's, Hb H, the embryonic Hb Portland I, Hb Portland II, Hb Portland III, and Hb Constant Spring (Hb CS) by HPLC using a weak ion

exchanger and the work of other investigators have encouraged the use of this technique to overcome false positive diagnosis of sickle cell disease sometimes found with traditional electrophoretic determinations². In spite of these advances, information on HPLC reference ranges is scarce or not readily available for the identification and quantification of hemoglobin variants.

Although it is recognized that HPLC reference ranges may vary from one laboratory to the next owing to materials, type of equipment and operating conditions employed, it is important that these parameters be established prior to its application in newborn/adult hemoglobin screening programs. The samples used in an high performance liquid chromatography machine is usually larger than that of a gas chromatography machine³. Despite the number of disadvantages it has, there are numerous advantages as well. The devices are more precise than other devices and can analyze a wider range of compounds, although the GC device can do better with more complex mixtures than the HPLC devices do. Our report deals with the development of normal hemoglobin reference range values for HPLC for a normal healthy adult population and the identification and quantification of several abnormal hemoglobins found in adult and newborn individuals⁴.

*Corresponding author: Gireesh. V. Achalkar

Department of Pathology, Raichur Institute of Medical Sciences, Raichur. Karnataka. 584102

There are more than 1,100 human hemoglobin variants⁵. The majority were discovered during population surveys and are not associated with clinical manifestations. OBJECTIVE: To establish normal baseline values for HB fractions.

MATERIALS AND METHODS

100 adults of age 18 to 50 and infants of 4 days to 300 days were selected for study. The clinical history and CBC findings were carefully studied. Subjects with normal CBC profile were included for study. Fifty microliter aliquots of fresh whole blood samples collected in ethylenediamine tetraacetic acid (EDTA) containing vacutainer tubes from normal newborn and adult individuals being screened at the were treated with a hemolyzate reagent centrifuged at 12,000 rpm at 4°C for 15 minutes and passed through a 0.25 to 0.45 diameter syringe filter unit into a 1.5 ml tube. In each case, 10ml of aliquot were transferred for hemoglobin variant analysis in a Hb Vario multisolvent HPLC system. The column used was a PolyCat A 300, stainless steel packed with hydrophilic cationic polymer; a gradient made up of mobile phase A containing 40 mM Bis Tris; 4 mM KCN at pH 6.5 and mobile phase B composed of 40 mM Bis tris; 4 mM KCN, 0.2 M NaCl at pH 6.8 and running conditions at 2.0 ml/min flow rate for 30 minutes, ultraviolet readings at 436 nm were observed. At the end of each run, qualitative and quantitative peak integration results were automatically obtained.

RESULTS

One hundred identified as normal adults (ages 18 to 50) by alkaline and acid electrophoresis and HPLC were chosen to establish the hemoglobin reference ranges. The corresponding hemoglobin ranges are shown in Table 1. The percentage values are as follows: Hb A mean of 92.3 percent ; HbA1 mean 3.6 percent ; Hb F mean of 3.2 percent ; Hb A2 mean of 2.1 percent , respectively.

Hemoglobin Percentages of Normal New borns and Babies

Hemoglobin percentile ranges for normal newborns and babies from 4 days up to 10 months of age were determined , and the hemoglobin trend changes occurring during this time period were also statistically assessed. The HPLC results registered in table II show that the percentage mean values and the minimum-maximum ranges for Hb F vary from 77.3 (64.6 to 90.1) at 4 days to 13.4 mean (8.0 to 18.0 %) at 300 days. TABLE 3 for Hb A vary from 23.1 range 9.9 to 36.4% at 4 days to 86.6 (81.2 to 92.0) at 300 days after birth. The decreasing trend for Hb F and the corresponding increments for Hb A observed after 4, 8 , 14, 20, 30, 50, 150, 210, and 300 days after birth are shown in Tables 2 and 3.

Table I High Performance Liquid Chromatography Hemoglobin Reference Ranges in Normal Adults

HB types	Mean %	SD	Minimum%	Maximum%
HBA	92.3	1.2	88.0	96.7
HBA1	3.6	0.6	1.0	6.2
HBF	3.2	0.6	1.2	5.1
HBA2	2.1	0.5	0.6	3.7

Table II High Performance Liquid Chromatography Hemoglobin F Percentage Trend Variations in Normal Newborns and Babies Up to 10 Months

Age in days	Mean	SD	Minimum	Maximum
4	77.3	7.6	64.6	90.1
8	75.8	8.2	60.5	91.2
14	76.1	8.6	57.9	94.4
20	78.0	5.3	68.0	88.0
30	74.2	7.1	62.2	86.2
60	52.8	3.3	32.1	73.6
150	27.3	2.9	21.1	33.5
210	26.1	2.9	21.0	31.2
300	13.4	5.0	8.0	18.8

Table III High Performance Liquid Chromatography Hemoglobin A Percentage Trend Variations in Normal Newborns and Babies Up to 10 Months

Age in days	Mean	SD	Minimum	Maximum
4	23.1	7.6	9.9	36.4
8	24.1	8.2	8.7	39.5
14	23.8	8.6	5.6	42.1
20	22.0	5.3	12.0	32.0
30	25.8	7.1	13.8	37.8
60	47.1	3.2	26.4	67.9
150	72.7	2.4	66.5	78.9
210	73.9	3.0	68.8	79.0
300	86.6	5.2	81.2	92.0

DISCUSSION

Current hemoglobin screening for the identification of each of the numerous hemoglob in abno rmalities known requires the use of at least two procedures to reach a definite diagnosis; with th e more difficult cases, analysis of deoxyribonucleic acid (DNA) is necessary.^{6,7} The use of cation exchange HPLC has added a tremendous value to hemoglobin variant detection for its ability to provide accurate qualitative data which is particularly important for the differential diagnosis of conditions such as SS and S(+ thal), CC and C(+ thal), and EE and E(+ thal) in newborns^{8,9}.

Because HPLC hemoglobin variant quantitative measurements vary significantly with respect to those compiled from other methodologies, ergo densitometric results from cellulose, citrate agar or isoelectrofocusing electrophoresis^{10,11}, the establishment of specific reference ranges to explore the quantitative capabilities of HPLC to differentiate the boundaries of a normal and abnormal hemoglobin conditions are being reported.¹²

High performance liquid chromatography hemoglobin ranges of Hb A, Hb A1; Hb F and Hb A2 encountered with a representative normal adult population fall within limits of normal controls derived from densitometric quantitation of cellulose acetate, citrate and/or isoelectrofocusing electrophoresis procedures^{13,14}; *i.e.*, the latter showing that the approximate mean percentage for each of these hemoglobins is: Hb A, 90 percent; Hb A1; 6.5 percent; Hb F, 1.0 percent; and Hb A2, 2.5 percent^{10,15,16}. The normal adult reference ranges shown in table 1 can be used with a certain degree of accuracy to detect the presence of an abnormal hemoglobin when the HPLC percentage results of a blood specimen does not correspond to these normal ranges. Hemoglobin statistical evaluation of normal newborns by HPLC varies with age, as expected^{17,18}. Newborn percentage trends shift from Hb F of 77.3 percent and Hb A of 23.1 percent at 4 days to Hb F mean of 13.4 and Hb A mean of 86.6 percent at 300 days after birth.

The reported HPLC normal newborn Hb F percentages fall within the 70 to 85 percent values obtained by other procedures^{19,20}; however, our HPLC results indicate that Hb F disappears in time at a much slower pace, while the Hb A percentage increments in the same fashion. In fact, after 150 days (5 months), 210 days (7 months), and 300 days (10 months), the mean percentage levels of Hb F were 27.3, 26.1, and 13.4, respectively. These results are in contrast with the Hb F rapid disappearing reports of 1 to 2 percent after 6 months^{13,21,22}. The apparent relatively high levels of Hb F (15.0 percent) after 10 months obtained by the HPLC method should be taken into consideration when trying to interpret Hb F percentages of babies one year of age or older.^{23,24,25}

Inherited abnormalities of hemoglobin synthesis may be due structural changes or due to reduced synthesis²⁶. Many are functionally normal and therefore clinically silent^{27,28}. Some form polymers (HBS) or crystals (HBC)^{29,30}. Abnormal Hemoglobins in Newborn and adult sickle cell trait individuals, Hb FAS and Hb AS, are readily identified by cation exchange HPLC^{12,31}. is in agreement with current hemoglobin detection protocols reported for adult Hb AS patients. However, when these same results were expressed in terms of Hb A/S ratios, Evaluation by HPLC of Hb FAS newborns at various age groups show a larger diminishing trend for Hb F and the corresponding increasing percentage of Hb A and Hb S with time when compared to normal babies, Hb FA or AA,. In all cases, the A/S ratios remain constant with a mean of 1.5 (s.d. range 1.1 to 2.2) regardless of age group^{32,33}. If this A/S ratio is compared with the adult Hb AS, there is practically no difference³⁴. These results may suggest that HB A/S ratios reported may indicate the presence of other type of hemoglobin inherited disorder which will require additional confirmatory tests³⁵, i.e., thalassemias; otherwise, the Hb AS trait condition of the patient would prevail.

The purpose of newborn hemoglobinopathy screening is to detect sickle cell disease³⁶. The most common types of sickle cell disease are sickle cell anemia (Hemoglobins SS), Hemoglobin SC disease, sickle beta thalassemia zero (Sβ⁰) and sickle beta thalassemia plus (Sβ⁺)^{37,38}. These conditions render infants susceptible to overwhelming pneumococcal infection and acute splenic sequestration^{39,40}. These life-threatening complications may occur prior to other less severe complications that would lead to the routine diagnosis and institution of preventive measures.⁴¹

CONCLUSION

The HPLC hemoglobin ranges for normal adult and newborns have been established. Application of these ranges to confront abnormal hemoglobins like Hb AS, Hb FAS, Hb AC, and HB FAC has shown that the A/X ratio pattern remains constant, even if it is a newborn. Detection of small amounts of Hb A, Hb S, or Hb C in newborns who have large quantities of Hb F has been possible by HPLC owing to its high sensitivity and specificity. Usefulness of this approach will be further attested with time when HPLC analysis of blood specimens for newborn screening and identification of hemoglobin variants becomes widespread in this country.

References

- Hattori Y, Kutlar F, Mosley CJ, et al. Association of the level of Gy chain in the fetal hemoglobin of normal adults with specific haplotypes. *Hemoglobin* 1986;10:185-204.
- Huisman THJ. Usefulness of cation exchange high performance liquid chromatography as a testing procedure. *Pediatrics* 1989;83(5 pt 2): 849-51.
- Ou CN, Buffone GJ, Reimer GL, Alpert AJ. High-performance liquid chromatography of human hemoglobins on a new cation exchanger. *J Chrom* 2 1983;266:197-205.
- Ou CN, Rognerud CL. Rapid analysis of hemoglobin variants by cation-exchange HPLC. *Clin Chem* 1993;39:820-4.
- Roa PD, Turner E, Aguinaga MdP. Hemoglobin variant detection from dried blood specimens by high performance liquid chromatography. *Ann Clin Lab Sci* 1993;23:433-8.
- Rogers BB, Wessels RA, Ou CN, Buffone GF. High-performance liquid chromatography in HEMOGLOBIN VARIANTS/HPLC/REFERENCE RANGES 235 the diagnosis of hemoglobinopathies and thalassemias. *Amer J Clin Path* 1985;84:67H.
- Wilson JB, Wrightstone RN, Huisman THJ. Rapid cation-exchange high-performance liquid chromatography procedure for the separation and quantitation of hemoglobins S, C, and O Arab in cord blood samples. *J Lab Clin Med* 1986; 108:138-41.
- Bellisario R, Barry RJ, Hamilton R, Pass KA. Newborn screening for hemoglobinopathies using a rapid automated membrane convective liquid chromatography system. *J Internat Soc Neonatal Screening* 1993;2:159-63.
- Inoue H, Takabe F, Maeno Y, Iwasa M. Identification of fetal hemoglobin in blood stains by high performance liquid chromatography. *Z Rechtsmed* 1989;102:437-44.
- Kutlar F, Gu LH, Hu H, Huisman THJ. Quantitation of hemoglobin's Bart's, H. Portland-I, Portland II and Constant Spring by anionexchange high performance liquid chromatography. *J Clin Chem* 1989;487:265-74.
- Kutlar F, Fei J, Wilson JB, Kutlar A, Huisman THJ. Detection of the embryonic chain in blood from newborn babies by reverse-phase high performance liquid chromatography. *J Chrom* 1987;394:333-43.
- Resolve-Hb kit analysis of adult and neonatal blood by isoelectric focusing electrophoresis. Isolab Inc., Akron, OH. Innovative Biochemical Methodology, February 1988.
- Huisman THJ, Schroeder WA. New aspects of the structure, function and synthesis of hemoglobin. *CRC Crit Rev Clin Lab Sci* 1970; 1:471-2.
- Weatherall DJ, Clegg JB. *The Thalassemia Syndromes*. Oxford, Blackwell Science, 2001.
- Sood SK, Madan N, Colah R, Sharma S, Apte SV eds Collaborative Study on thalassemia - An ICMR Task Force Study, New Delhi. *Indian Council of Medical Research* 1993.
- Desai SN, Colah RB. Alpha thalassemia syndromes in India. *Ind J Hum Genet* 1997; 3: 1-9.
- Clarke GM, Higgins TN. Laboratory investigations of hemoglobinopathies and thalassemias: Review and update. *Clin Chem* 2000; 46: 1284-1290.
- Bild BJ, Bain BJ. Investigation of abnormal hemoglobins and thalassemia. In Lewis SM, Bain BJ, Bates I, eds. *Dacie & Lewis Practical Hematology*. 9th ed. Churchill Livingstone 2001; 231-268.
- George E, Khoo SK, Mukhtar AB, Nor Aini U. Screening for thalassemia in pregnant women: A

- laboratory perspective. *Malaysian J Med & Health Sciences* 2005; 1: 111-117.
20. Working Party of the General Hematology Task Force of the British Committee for Standards in Hematology. Guideline: The laboratory diagnosis of hemoglobinopathies. *Brit J Hematol* 1998; 101: 783-792.
 21. Jones W and Clark LA. Hemoglobinopathy and thalassemia testing: An automated method. *Amer. Clin. Lab.* Nov 1994.
 22. Bain BJ. *Hemoglobinopathy diagnosis*. England; Oxford, Blackwell Science Ltd, 2001; 260.
 23. Wajcman H, Prehu C, Bardakdijian-Michau J, Prome D, Riou J, Godart C, Mathis M, Hurtrel D, Galacteros F. Abnormal hemoglobins: Laboratory methods. *Hemoglobin* 2001; 25: 169-181.
 24. Mario N, Baudin B, Aussel C, Giboudeau J. Capillary IEF and HPLC compared for the qualitative and quantitative analysis of hemoglobin variants. *Clin Chem* 1997; 43: 2137-2142.
 25. Suh DD, Kruss JS, Bures K. Influence of Hb S adducts on HbA₂ quantitation by HPLC. *Clin Chem* 1996; 42: 113-114.
 26. Cotton F, Gulbis B, Hansen V, Vertongen F. Interference of Hb D in HbA₂ measurement by cation exchange HPLC. *Clin Chem* 1999; 45: 1317-1318.
 27. Joutovsky A, Hadzi-Nesic J, Nardi MA. Retention time as a diagnostic tool for hemoglobin variants and hemoglobinopathies: A study of 60,000 samples in a clinical diagnostic laboratory. *Clin Chem* 2004; 50: 1736-1747.
 28. Colah RB, Wadia M, Surve R, Nadkarni A, Phanasaonkar S, Gorakshakar A, Mohanty D, Prome D, Wajcman H. Hb D Agri [β 9 (A6) Ser \rightarrow Tyr, β 121 (GH - 4) Glu \rightarrow Gln]: A new Indian hemoglobin variant with two amino acid substitutions in the same beta chain. *Hemoglobin* 2001; 25: 317-321.
 29. Wajcman H, Riou J, Yapo AP. Globin chain analysis by reversed phase high performance liquid chromatography: Recent developments. *Hemoglobin* 2002; 26: 271-284.
 30. Henthorn JS, Almeida AM, Davies SC. Neonatal screening for sickle cell disorders. *Brit J Hematol* 2004; 124: 259-263.
 31. Campbell M, Henthorn JS, Davies SC. Evaluation of cation exchange HPLC compared with isoelectric focusing for neonatal hemoglobinopathy screening. *Clin Chem* 1999; 45: 969-975.
 32. Panadea C, Eckman JR, Kuchnert RS, Platt AF. Comparison of liquid and dried blood for neonatal hemoglobin screening: Laboratory and programmatic issues. *Pediatrics* 1994; 93: 427-432.
 33. Angastiniotis M, Modell B, Boulinzhenkov V. Prevention and control of hemoglobinopathies. *Bull WHO* 1995; 73: 375-386.
 34. Samawat A, Modell B. Iranian national thalassemia screening programme. *Brit Med J* 2004; 329: 1134-1137.
 35. Wadia MR, Phanasaonkar SP, Nadkarni AH, Surve RR, Gorakshakar AC, Colah RB, Mohanty D. Usefulness of automated chromatography for rapid fetal blood analysis for second trimester prenatal diagnosis of β -thalassemia. *Prenatal diagnosis* 2002; 22: 153-157.
 36. Winichagoon P, Sriphanich R, Sae-Ngow B, Chowthaworm J, Tantisirin P, Kanokpongsakdi S, Fucharoen S, Wasi P. Application of automated HPLC in prenatal diagnosis of thalassemia. *Lab Hematol* 2002; 8: 29-35.
 37. Colah R, Surve R, Nadkarni A, Gorakshakar A, Phanasaonkar S, Satoskar P, Mohanty D. Prenatal diagnosis of sickle syndromes in India: dilemmas in counseling. *Prenatal Diagnosis* 2005; 25: 345-349.
 38. Colosimo A, Guida V, De Luca A, Coppabianca MP, Biancoi I, Palka G, Dallapiccola B. Reliability of DHPLC in mutational screening of β globin (HBB) allele. *Hum Mutat* 2002; 19: 287-295.
 39. Wu G, Hua I, Zhu J, Mo QH, Xu XM. Rapid accurate genotyping of β -thalassemia mutations using a novel multiplex primer extension/denaturing high performance liquid chromatography assay. *Brit J Hematol* 2003; 122: 311-316.
 40. Guida V, Colosimo A, Fichera M, Lombardo T, Rigoli L, Dallapiccola B. Hematologic and molecular characterization of a Sicilian cohort of α thalassemia carriers. *Haematologica* 2006; 91: 409-410.
 41. Colah RB, Gorakshakar AC. Thalassemias: Molecular genetics in antenatal diagnosis. In Lokeshwar MR, Marwaha RK, Shah M. eds. *Recent advances in Pediatrics*. New Delhi; Jaypee Brothers, 2000: 22-45.

How to cite this article:

Gireesh. V. Achalkar (2018) 'Hemoglobin Fractionation By HPLC', *International Journal of Current Medical And Pharmaceutical Research*, 04(8), pp. 3600-3603.
